# Utility of Serum Paraoxonase Levels with reference to Severity of Organophosphorus Poisoning

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## **ABSTRACT**

Introduction: Organophosphorus (OP) compounds are widely used insecticides for agricultural and domestic purposes. Easy availability and less awareness regarding the toxicity caused by these compounds have resulted in high morbidity and mortality in India. Early diagnosis and initiation of treatment are required to reduce the mortality rate for which laboratory evaluation plays a vital role, in addition to various clinical scoring systems.

**Materials and methods:** A cross-sectional study was carried out for a period of 2 months. Forty clinically diagnosed acute OP poisoning cases admitted in emergency units formed the study subjects. Serum was used for the estimation of cholinesterase, for both basal and salt stimulated paraoxonase (PON) activity. Peradeniya organophosphorus poisoning (POP) scale was used as a tool to categorize patients into mild (0–3 score), moderate (4–7 score), and severe (8–11 score) poisoning.

**Results:** The mean age of the study participants was  $31.9 \pm 14.4$  years. Seventy-five percent of the participants were males and 25% were females. Chlorpyrifos was the most common OP compound consumed by the study participants. There was a significant decrease in the serum cholinesterase activity (p = 0.001) and salt-stimulated PON activity (p = 0.016) as the severity increased. Serum cholinesterase and POP score showed statistically significant negative correlation (p = 0.003). There was a linear positive correlation between serum cholinesterase and serum PON activity, but the correlation was significant only with salt-stimulated PON activity (p = 0.005).

**Conclusion:** The results suggest that subjects with higher levels of PON activity may have better detoxifying capacity toward OP poisoning.

**Keywords:** Acute toxicity, Butyrylcholinesterase, Chlorpyrifos, Organophosphorus, Paraoxonase 1.

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#### INTRODUCTION

A national survey in India reported suicide as the cause for 3% surveyed deaths and about half of these deaths were due to poisoning mainly with ingestion of pesticides. Organophosphorus compounds are the widely used insecticides in agricultural industry. Easy availability and less awareness regarding the toxicity caused by these compounds have resulted in high morbidity and mortality in India. The OP compounds act by inhibiting the acetyl cholinesterase enzyme at muscarinic and nicotinic receptors, producing an array of symptoms like miosis, bradycardia, increased gastrointestinal motility, emesis, sweating, etc.<sup>2</sup>

The OP compounds are biotransformed *in vivo* through oxidative desulfuration and dealkylation to form its oxygen analogs and other active metabolites. These metabolites are potent inhibitors of the enzyme acetyl cholinesterase.<sup>3</sup> Detoxification of these active metabolites predominantly occurs through hydrolysis catalyzed by PON enzyme.<sup>4</sup> Early diagnosis and initiation of treatment are required to reduce mortality rate for which laboratory evaluation plays a vital role. In addition, various clinical scoring systems are available. Until now, measurement of serum cholinesterase has been considered as the most specific test for OP poisoning.<sup>5</sup> Studies in animal models have demonstrated high-density lipoprotein-associated paraoxonase (PON1) as one of the important determinants of an individual's sensitivity to some OP insecticides.<sup>6-8</sup>

Despite having convincing evidence in animal models, very few epidemiological studies have examined PON status as a determinant of severity of acute OP poisoning. The present study was aimed to examine the utility of serum PON levels with reference to severity of the poisoning in humans with acute exposure to OP compounds and correlate the PON activity with serum cholinesterase levels and POP scale.

## **MATERIALS AND METHODS**

A cross-sectional study was carried out at a rural tertiary care teaching college and hospital for a period of 2 months. Forty acute OP poisoning cases admitted in emergency unit with diagnosis confirmed by circumstantial evidence, history, and clinical examinations were included in the study, and formed the "convenience sample." Study

Table 1: Age-wise distribution of study participants

Age in years	Number (%)
0–20	5 (12.5)
21–40	28 (70)
Above 40	7 (17.5)
Total	40 (100)

subjects of either gender in the age group 18 to 50 years were included. Patients with history of other poisoning, psychiatric disorders, pregnancy, and other illness, such as chronic renal failure, diabetes mellitus, epilepsy, and myopathy were excluded from the study.

Under aseptic precautions, 5 mL of blood was drawn at the time of presentation from all the study subjects. Serum was used for the estimation of cholinesterase, for both basal and salt stimulated paraoxonase (PON) activity. Cholinesterase activity in serum was estimated by kinetic colorimetric method using Agappe liquichek cholinesterase kit, which is based on new Deutsche Gesellschaft Fur Klinische Chemie recommendations.<sup>9,10</sup> The PON activity was determined spectrophotometrically by using p-nitro phenyl acetate as a substrate. 11,12 The POP scale 13 was used for categorizing patients into mild (0-3 score), moderate (4-7 score), and severe (8-11 score) poisoning. An informed consent was taken from all participants and all the experiments involving human subjects were carried out in accordance with the protocol approved by Institutional Ethical Committee.

# **Statistical Analysis**

Data were entered into the excel sheet. The entire variables were tabulated, and mean and standard deviation (SD) were calculated for age, serum cholinesterase, and PON. Comparison of means between the groups was done using analysis of variance (ANOVA). Correlation analysis between serum PON levels with serum cholinesterase and severity of the disease was done using Pearson's correlation coefficient. The p-value less than 0.05 was considered statistically significant. All statistical analysis was done using Statistical Package for the Social Sciences software version 18.

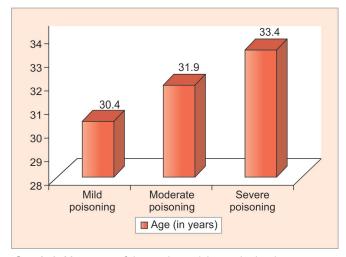
## **RESULTS**

Serum cholinesterase and PON activity were determined in the study participants, who had acute exposure to OP compounds. Majority (70%) of the study participants belonged to 21 to 40 years age group (Table 1). The mean age of the study participants was  $31.9 \pm 14.42$  years. 75% (n = 30) of the participants were males and 25% (n = 10) were females. The mean serum cholinesterase levels among subjects was  $4709 \pm 2829.69$  U/L and mean serum basal and salt-stimulated PON

**Table 2:** Demographic profile and mean values of serum cholinesterase and serum PON activity in OP poisoning cases

Variables		Total study participants mean ± SD
Age in years		31.9 ± 14.4
Sex	Male [N (%)]	30 (75)
	Female [N (%)]	10 (25)
Serum cholinesterase (U/L)		4709.18 ± 2829.69
Paraoxonase activity	Basal activity	55.33 ± 12.42
(nmol/mL/minute)	Salt-stimulated activity	48.68 ± 16.46

SD: Standard deviation



Graph 1: Mean age of the study participants in the three groups

levels were  $55.33 \pm 12.42 \text{ nmol/mL/minute}$  and  $48.68 \pm 16.46 \text{ nmol/mL/minute}$  respectively (Table 2). The mean age was higher in severe poisoning group when compared with other two groups mild and moderate poisoning group (Graph 1). Chlorpyrifos was the most common OP compound consumed (45%) followed by Malathion (22.5%), Diazinon (7.5%), fention (7.5%), Dichlorvos (5%), and other OP compounds (12.5%); (Table 3). There was statistically significant decrease in the serum cholinesterase activity (p = 0.001) and salt-stimulated PON activity (p = 0.016) as the severity of disease increased (Table 4 shows results of ANOVA). Serum cholinesterase and POP score showed negative correlation and was statistically significant (p = 0.0036).

**Table 3:** Details of OP compound consumed by the study participants

Name of the OP compound	Number (%)
Chlorpyrifos	18 (45)
Malathion	9 (22.5)
Diazinon	3 (7.5)
Fenthion	3 (7.5)
Dichlorvos	2 (5)
Other OP compounds	5 (12.5)
Total	40 (100)



Table 4: Serum cholinesterase and serum PON activity in different groups of OP poisoning cases

	Mild poisoning	Moderate poisoning	Severe poisoning	
Measures	(n = 13)	(n = 15)	(n = 12)	p-value
Serum cholinesterase (U/L				
Mean ± SD	7959.85 ± 1127.67	4642.53 ± 1045.10	1270.92 ± 470.16	0.001*
Median	7543	4848	1246	
Range	6388–9831	3036–6094	589-1948	
Serum Paraoxonase activ	ity:basal (nmol/mL/minute)			
Mean ± SD	$59.60 \pm 7.77$	53.09 ± 12.82	53.50 ± 15.52	0.32
Median	59.62	57.94	56.67	
Range	46.64-69.34	24.94-66.31	17.56-76.08	
Serum Paraoxonase activ	ity: salt stimulated (nmol/mL/minute)			
Mean ± SD	54.11 ± 13.25	52.78 ± 13.34	37.66 ± 18.78	0.016*
Median	59.93	54.12	31.61	
Range	31.99–69.43	17.56-68.24	16.46-67.76	

<sup>\*</sup>Statistically significant; SD: Standard deviation

**Table 5:** Pearson correlation of serum cholinesterase, serum PON, and POP score

Parameter	r-value	p-value
Serum cholinesterase with POP score	-0.449	0.003*
Serum cholinesterase with basal paraoxonase activity	0.234	0.146
Serum cholinesterase with salt-stimulated paraoxonase activity	0.429	0.0057*
Basal paraoxonase activity with POP score	-0.282	0.077
Salt-stimulated paraoxonase activity with POP score	-0.1521	0.349

<sup>\*</sup>Statistically significant

There was a linear positive correlation between serum cholinesterase activity and serum PON among the study participants, but the correlation was significant only with salt-stimulated PON activity (p = 0.0057). Even though there was negative correlation between POP score and PON activity among subjects, it was not statistically significant (Table 5).

# DISCUSSION

The OP compounds inhibit acetylcholinesterase activity at the nerve endings resulting in an array of signs and symptoms mainly due to muscarinic and nicotinic receptor overstimulation. Although estimation of acetylcholinesterase activity is more specific, estimation of plasma cholinesterase also known as butyrylcholinesterase or psuedocholinesterase is done as a surrogate for neuronal acetylcholinesterase. In acute exposure to OP compounds, the severity of the poisoning parallels the decrease in psuedocholinesterase activity.<sup>14</sup>

In the present study, the serum cholinesterase level was low in severe poisoning group when compared with mild poisoning and moderate poisoning groups. The decrease in this enzyme level was statistically significant (p = 0.001) and is in accordance with few other

studies done in South India in subjects exposed to acute OP compounds toxicity. 14,15

The quest for newer and specific biomarkers in relation to OP poisoning started quite a long time back. In this regard, many studies have proposed that measurement of PON status is a reliable marker to assess susceptibility to OP poisoning. <sup>16,17</sup>

In many animal models, association between PON1 status and severity or susceptibility to OP poisoning has been well characterized. <sup>6,7,16</sup> Several studies in individuals with chronic exposure to OP compounds have demonstrated high risk of OP toxicity and low levels of PON enzyme activity. <sup>18,19</sup>

However, there are very few epidemiological studies done in humans to explore the PON status in acutely exposed OP poisoning individuals. In the present study, the levels of both basal and salt-stimulated PON activity decreased from mild to severe poisoning group, but this decrease was statistically significant only with salt-stimulated PON activity. This difference in the salt stimulation can be attributed to the large variation of genetically determined serum PON1 activity in the human population. Also Akgur et al<sup>20</sup> proposed that individuals with low PON1 activity on salt stimulation also had low cholinesterase activity and are more susceptible to OP poisoning. Ceron et al<sup>21</sup> in their review have reported that when NaCl was used with paraoxon as substrate for phenotype characterization, it increases the PON1 activity, but when used with other substrates, such as phenyl acetate, it produces a decrease in PON1 activity.

Animal models have demonstrated that the animals with low levels of PON enzyme activity in their sera are very much susceptible to OP poisoning,<sup>22-24</sup> and by increasing the levels of this enzyme by intravenous injection of partially purified PON1, it has thrown some light on the protective role of PON against OP toxicity.<sup>25,26</sup>

Studies done on birds have shown that low PON1 activity displayed higher sensitivity to OP.<sup>27</sup> Further studies<sup>22,23</sup> have demonstrated that administration of exogenous purified PON1 has increased the resistance to the toxic effects of OPs among mice and rats and also offered protection against toxic effects of Chlorpyrifos when given within 3 hours of OP exposure.<sup>26</sup>

Recently, it has been projected that administration of recombinant human PON1 to mice has resulted in protection against the toxicity of Chlorpyrifos. <sup>28</sup> All these studies indicate that by artificially modulating the levels of PON1 in serum, it is possible to decrease the toxicity of certain OP compounds. Series of research done on genetically modified mice for the expression on human PON1 genes and its effect on OP toxicity <sup>6,7</sup> has provided convincing evidence for extrapolating these results to humans.

According to the present study, results of PON activity, both basal and salt stimulated, showed linear positive correlation with serum cholinesterase levels, but was statistically significant with only salt-stimulated activity (p = 0.0057). This result indicates the association between serum PON and serum cholinesterase and is consistent with the results of few others,  $^{15,18,29}$  but not all.  $^{30,31}$ 

These conflicting results may be due to difference in the study population, type, and duration of exposure and presence of PON gene polymorphism. The most common PON1 gene polymorphism due to substitution at position 192 (PON1 $_{Q192R}$ ) directly affects its hydrolytic efficiency. The alloform PON1 $_{QQ192}$  detoxifies Chlorpyrifos and diazoxin more efficiently than paraoxon, which is rapidly hydrolyzed by PON1 $_{RR192}$  alloform.  $^{32,33}$ 

Individuals with low serum cholinesterase activity also had low levels of PON activity belonging to severe poisoning group suggesting that these individuals are more susceptible to OP poisoning than those with higher levels of PON and serum cholinesterase activity. This indicates that hydrolysis of OP compound by PON is a major factor determining the toxicity. Hence, estimation of serum PON along with serum cholinesterase might throw some light in predicting and assessing the detoxifying capacity of patients, thereby, aiding in treatment protocol of patients with acute exposure to OP compound.

## **LIMITATIONS**

- The present study had low sample size and adopted convenient sampling technique.
- The subjects were not followed up for outcome of the toxicity.
- The other confounding factors like drugs (statins), history of any chronic exposure to OP compounds, etc., were not considered.

## CONCLUSION

The present study revealed that estimation of serum PON activity determines the OP toxicity; thereby, it helps in predicting the severity of OP poisoning. The PON is proved to be a good surrogate for cholinesterase activity in serum. The results suggest that the patients with higher levels of PON activity may have substantially better detoxifying capacity against OP compounds. However, the direct and conclusive confirmation in human of the relevance of PON1 status in determining relative sensitivity to OP toxicity is still lacking. Further research can be undertaken to investigate the same in future.

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## **REFERENCES**

- Patel V, Ramasundarahettige C, Vijayakumar L, Thakur JS, Gajalakshmi V, Gururaj G, Suraweera W, Jha P, Million Death Study Collaborators. Suicide mortality in India: a nationally representative survey. Lancet 2012 Jun 23;379(9834): 2343-2351.
- 2. Bhattacharyya K, Phaujdar S, Sarkar R, Mullick OS. Serum creatine phosphokinase: a probable marker of severity in organophosphorus poisoning. Toxicol Int 2011 Jul;18(2): 117-123.
- Elersek T, Filipic M. Organophosphorus pesticide-mechanism of their toxicity. In:Stoyteheva M, editors. Pesticide – The impacts of pesticides exposure. 1st ed. Rijeka: In Tech; 2011. p. 243-260.
- Costa LG, Richter RJ, Li WF, Cole T, Guizzetti M, Furlong CE. Paraoxonase (PON 1) as a biomarker of susceptibility for organophosphate toxicity. Biomarkers 2003 Jan-Feb;8(1):1-12.
- Thiermann H, Kehe K, Steinritz D, Mikler J, Hill I, Zilker T, Eyer P, Worek F. Red blood cell acetylcholinesterase and plasma butyrylcholinesterase status: important indicators for the treatment of patients poisoned by organophosphorus compounds. Arh Hig Rada Toksikol 2007 Sep;58(3):359-366.
- Cole TB, Walter BJ, Shih DM, Tward AD, Lusis AJ, Timchalk C, Richter RJ, Costa LG, Furlong CE. Toxicity of chlorpyrifos and chlorpyrifos oxon in a transgenic mouse model of the human paraoxonase (PON1) Q192R polymorphism. Pharmacogenet Genomics 2005 Aug;15(8):589-598.
- Li WF, Costa LG, Richter RJ, Hagen T, Shih DM, Tward A, Lusis AJ, Furlong CE. Catalytic efficiency determines the in-vivo efficacy of PON1 for detoxifying organophosphorus compounds. Pharmacogenetics 2000 Dec;10(9):767-779.
- Shih DM, Gu L, Xia YR, Navab M, Li WF, Hama S, Castellani LW, Furlong CE, Costa LG, Fogelman AM, et al. Mice lacking serum paraoxonase are susceptible to organophosphate toxicity and atherosclerosis. Nature 1998 Jul 16;394(6690):284-287.
- Recommendations of the German Society for Clinical Chemistry. Standardization of methods for the estimation of enzyme activities in biological fluids: Standard method for the determination of Cholinesterase activity. J Clin Chem Clin Biochem 1992;30:163-170.



- Panteghini M, Bais R, van Solinge WW. Enzymes. In: Burtis CA, Ashwood ER, Bruns DE, editors. Tietz Text book of Clinical chemistry and molecular diagnostics. 4th ed. New Delhi: Elsevier; 2008. pp. 597-644.
- 11. Nagane NS, Ganu JV. Lipid profile and serum paraoxonase1 activity in CRF patient's pre and post hemodialysis. Al Ameen J Med Sci 2011;4(1):61-68.
- 12. Charlton-Menys V, Liu Y, Durrington PN. Semiautomated method for determination of serum paraoxonase activity using paraoxon as substrate. Clin Chem 2006 Mar;52(3):453-457.
- Senanayake N, de Silva HJ, Karalliedde L. A scale to assess severity in organophosphorus intoxication: POP scale. Hum Exp Toxicol 1993 Jul;12(4):297-299.
- Sumathi ME, Kumar SH, Shashidhar KN, Takkalaki N. Prognostic significance of various biochemical parameters in acute organophosphorus poisoning. Toxicol Int 2014 May;21(2): 167-171.
- 15. Richard SA, Frank EA, D'Souza CJ. Correlation between cholinesterase and paraoxonase 1 activities: case series of pesticide poisoning subjects. Bioimpacts 2013;3(3):119-122.
- Costa LG, Cole TB, Vitalone A, Furlong CE. Measurement of paraoxonase (PON1) status as a potential biomarker of susceptibility to organophosphate toxicity. Clin Chim Act 2005 Feb;352(1-2):37-47.
- Sirivarasai J, Kaojarern S, Yoovathaworn K, Sura T. Paraoxonase (PON1) polymorphism and activity as the determinants of sensitivity to organophosphates in human subjects. Chem Biol Interact 2007 Jul 20;168(3):184-192.
- Hofmann JN, Keifer MC, Furlong CE, De Roos AJ, Farin FM, Fenske RA, van Belle G, Checkoway H. Serum cholinesterase inhibition in relation to paraoxonase-1 (PON1) status among organophosphate-exposed agricultural pesticide handlers. Environ Health Perspect 2009 Sep;117(9):1402-1408.
- 19. Mackness B, Durrington P, Povey A, Thomson S, Dippnall M, Mackness M, Smith T, Cherry N. Paraoxonase and susceptibility to organophosphorus poisoning in farmers dipping sheep. Pharmacogenetics 2003 Feb;13(2):81-88.
- Akgur SA, Ozturk P, Sozmen YE, Delen Y, Tanyalcin T, Ege B. Paraoxonase and acetylcholinesterase activities in human exposed to organophosphorus compounds. J Toxicol Environ Health A 1999 Dec 24;58(8):469-474.
- Ceron JJ, Tecles F, Tvarijonaviciute A. Serum paraoxonase1 (PON1) measurement: an update. BMC Vet Res 2014 Mar 25;10:74.
- 22. Costa LG, McDonald BE, Murphy SD, Omenn GS, Richter RJ, Motulsky AG, Furlong CE. Serum paraoxonase and its influence on paraoxon and chlorpyrifosox on toxicity in rats. Toxicol Appl Pharmacol 1990 Mar 15;103(1):66-76.

- Li WF, Costa LG, Furlong CE. Serum paraoxonase status: a major factor in determining resistance to organophosphates. J Toxicol Environ Health 1993 Oct-Nov;40(2-3):337-346.
- 24. Walker CH, Mackness MI. "A" esterases and their role in regulating the toxicity of organophosphates. Arch Toxicol 1987;60(1-3):30-33.
- 25. Main AR. The role of A-esterase in the acute toxicity of paraoxon, TEPP and parathion. Can J Biochem Physiol 1956 Mar;34(2):197-216.
- Li WF, Furlong CE, Costa LG. Paraoxonase protects against chlorpyrifos toxicity in mice. Toxicol Lett 1995 Apr;76(3): 219-226.
- 27. Brealey CB, Walker CH, Baldwin BC. A-esterase activities in relation to the differential toxicity of pirimiphos-methyl to birds and mammals. Pest Sci 1980;11:546-554.
- Cowan J, Sinton CM, Varley AW, Wians FH, Haley RW, Munford RS. Gene therapy to prevent organophosphate intoxication. Toxicol Appl Pharmacol 2001 May 15;173(1):1-6.
- 29. Goel P, Goel K, Singh S, Bhalla A, Sharma N, Gill KD, Bandyopadhyay D. Role of paraoxonases in detoxification of organophosphates. JARBS 2012;4(4):320-325.
- 30. Pérez-Herrera N, Polanco-Minaya H, Salazar-Arredondo E, Solís-Heredia MJ, Hernández-Ochoa I, Rojas-García E, Alvarado-Mejía J, Borja-Aburto VH, Quintanilla-Vega B. PON1Q192R genetic polymorphism modifies organophosphorous pesticide effects on semen quality and DNA integrity in agricultural workers from southern Mexico. Toxicol Appl Pharmacol 2008 Jul 15;230(2):261-268.
- 31. Ellison CA, Crane AL, Bonner MR, Knaak JB, Browne RW, Lein PJ, Olson JR. PON1 status does not influence cholinesterase activity in Egyptian agricultural workers exposed to chlorpyrifos. Toxicol Appl Pharmacol 2012 Dec 15;265(3): 308-315.
- 32. Kanamori-Kataoka M, Seto Y. Paraoxonase activity against nerve gases measured by capillary electrophoresis and characterization of human serum paraoxonase (PON-1) polymorphism in the coding region (Q192R). Anal Biochem 2009;385(1):94-100.
- 33. López-Flores I, Lacasaña M, Blanco-Muñoz J, Aguilar-Garduño C, Sanchez-Villegas P, Pérez-Méndez OA, Gamboa-Avila R. Relationship between human paraoxonase-1 activity and PON-1 polymorphisms in Mexican workers exposed to organophosphate pesticides. Toxicol Lett 2009 Jul 24;188(2): 84-90.
- 34. Mackness B, Mackness MI, Arrol S, Turkie W, Durrington PM. Effect of molecular polymorphisms of serum paraoxonase (PON1) on the rate of hydrolysis of paraoxon. Br J Pharmacol 1997 Sep;122(2):265-268.